

# BioMEMS (and Microfluidics)





Microfluidics & Implantable sensors



**BioMEMS**

**'Traditional' MEMS**

Accelerometers & Inject Heads

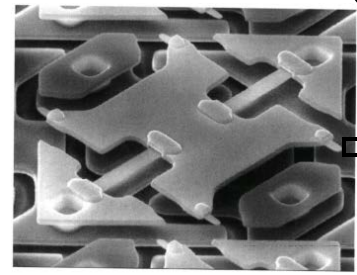
**MEMS Technology**

**RF MEMS**

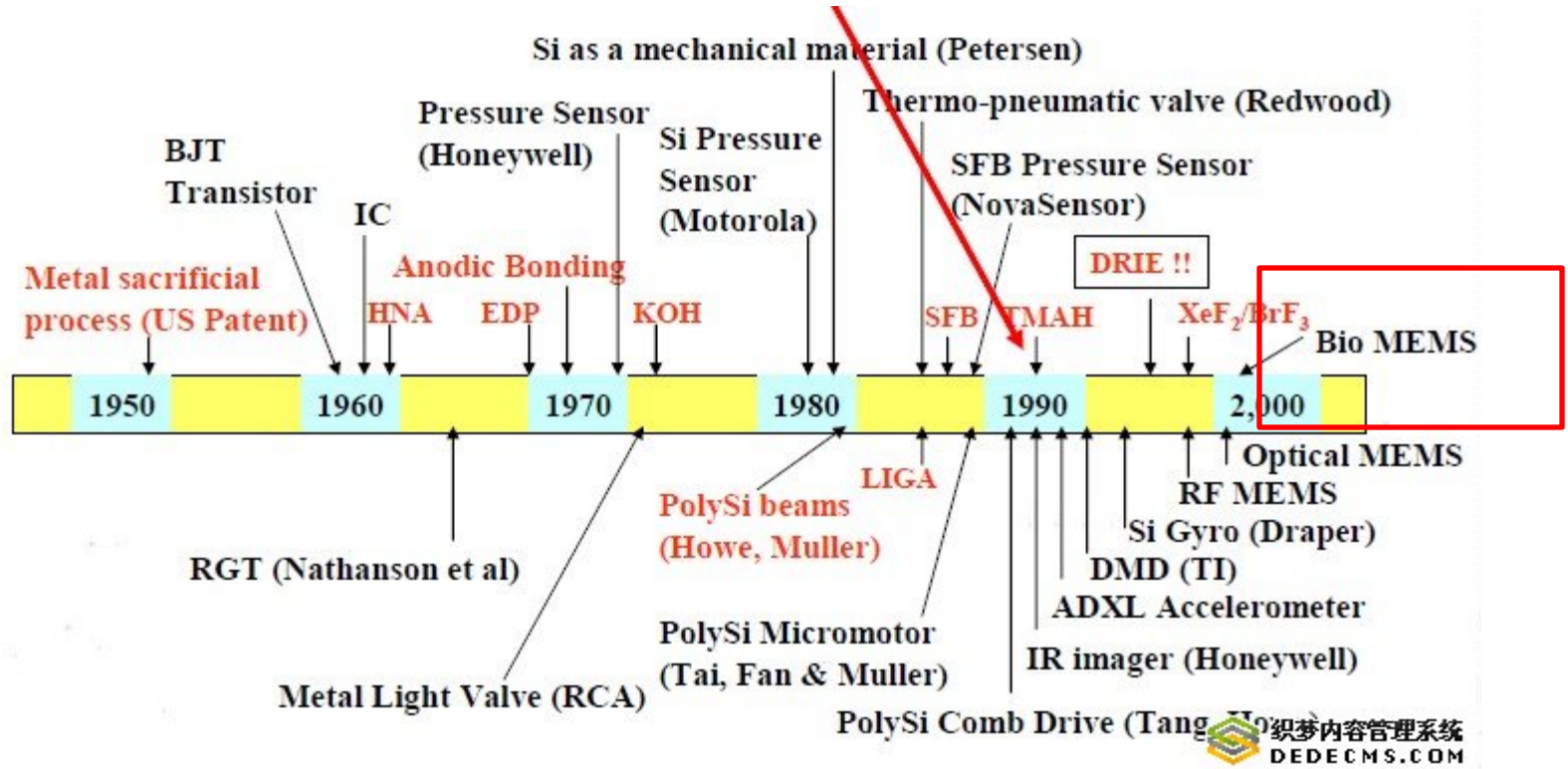
Filters & Varactors

**Optical MEMS (MOEMS)**

Mirrors & Switches



# History of MEMS Technology



**BioMEMS is a relatively new field...**

## SILICON & ITS DERIVATIVES

Silicon(Si)

Silicon Dioxide -  $\text{SiO}_2$  (glass)

Silicon nitride ( $\text{Si}_x\text{N}_y$ )

....



- Mechanical Reliability
- Performance
- IC compatibility

## POLYMERS

Photosensitive Polymers  
(e.g. SU-8)

Polydimethylsiloxane  
(PDMS)

Parylene

PS

PMMA

...



- Biocompatibility
- Cost
- Surface Modification
- Disposability (e.g. single use devices)
- Rapid Prototyping

## BioMEMS Materials

## METALS

Platinum

Silver

Chrome and Gold

Indium Tin Oxide (ITO)

...



- Increased Functionality
- Integration (sensors & actuators)

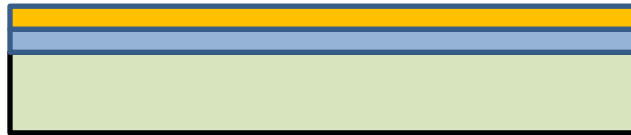
# Microfabrication Consists of 3 Major Steps: Deposition, Patterning, Removal



1. Select a Substrate (e.g. a silicon wafer)



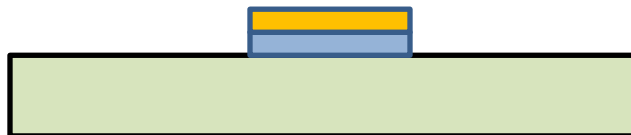
2. DEPOSIT the *Structural* Material (usually a few-microns thick film)



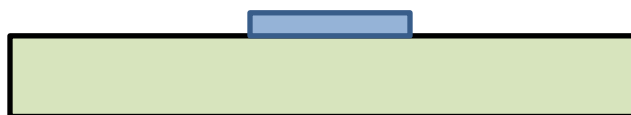
3. DEPOSIT PhotoResist (PR) (PR is photosensitive to UV radiation)



4. PATTERN PR using light (LITHOGRAPHY)

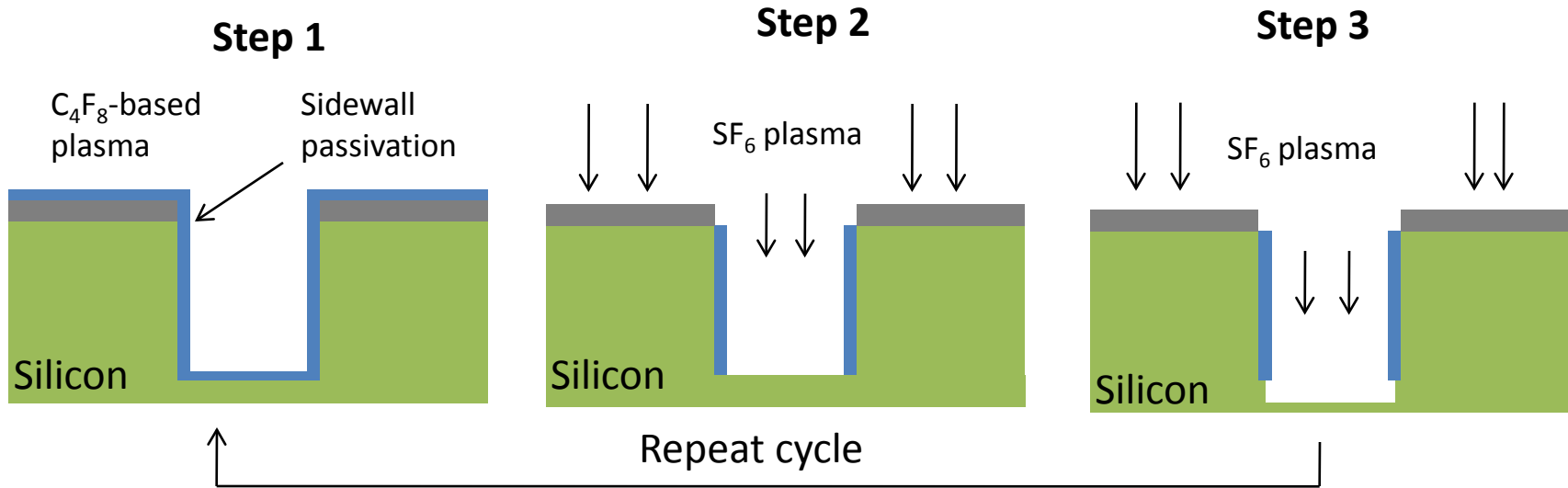


5. REMOVE the structural material



6. REMOVE PR

# DRIE of Si – Operation Principle



**Etching is performed in cycles of 3 steps:**

**Deposit Polymer (step 1) :**  $C_4F_8$ -based plasma is used to conformally deposit a few monolayers of PTFE-like fluorocarbon polymer across all surfaces

**Etch polymer (step 2):** The plasma gas is then switched to  $SF_6$  that isotropically etches silicon (like typical RIE). Ions from the plasma bombard the surface of the wafer, removing the polymer. Increased ion energy in the vertical direction results in a much higher rate of removal of fluorocarbon from surfaces parallel to the wafer surface.

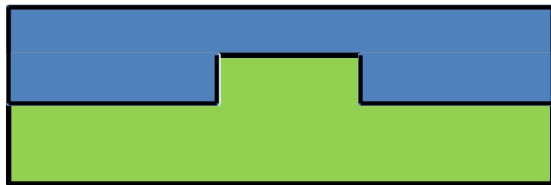
**Etch silicon (step 3) :** Following selective polymer removal, the silicon surface at the base of the trench is exposed to reactive fluorine-based species that isotropically etch the unprotected silicon. The remaining fluorocarbon polymer protects the vertical walls of the trench from etching.

# Soft- Lithography: Creating a 'Soft' (e.g. PDMS) Mold

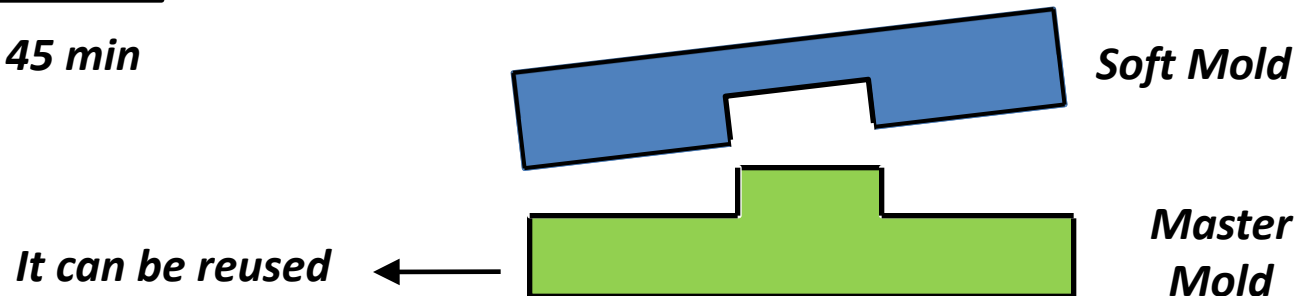
1. Start with a Master Mold



2. Cast and Cure PMDS



3. Peel off and u r DONE!



# What can you do with the 'Soft' Mold?

1. Micro Contact Printing ( $\mu$ CP)

2. Replica molding (REM)

**SOFT LITHOGRAPHY**

```
graph TD; A[SOFT LITHOGRAPHY] --> B[1. Micro Contact Printing (μCP)]; A --> C[2. Replica molding (REM)]; A --> D[3. Micromolding in capillaries (MIMIC)]; A --> E[4. Microtransfer molding (μTM)]; A --> F[5. Microfluidics];
```

3. Micromolding in capillaries  
(MIMIC)

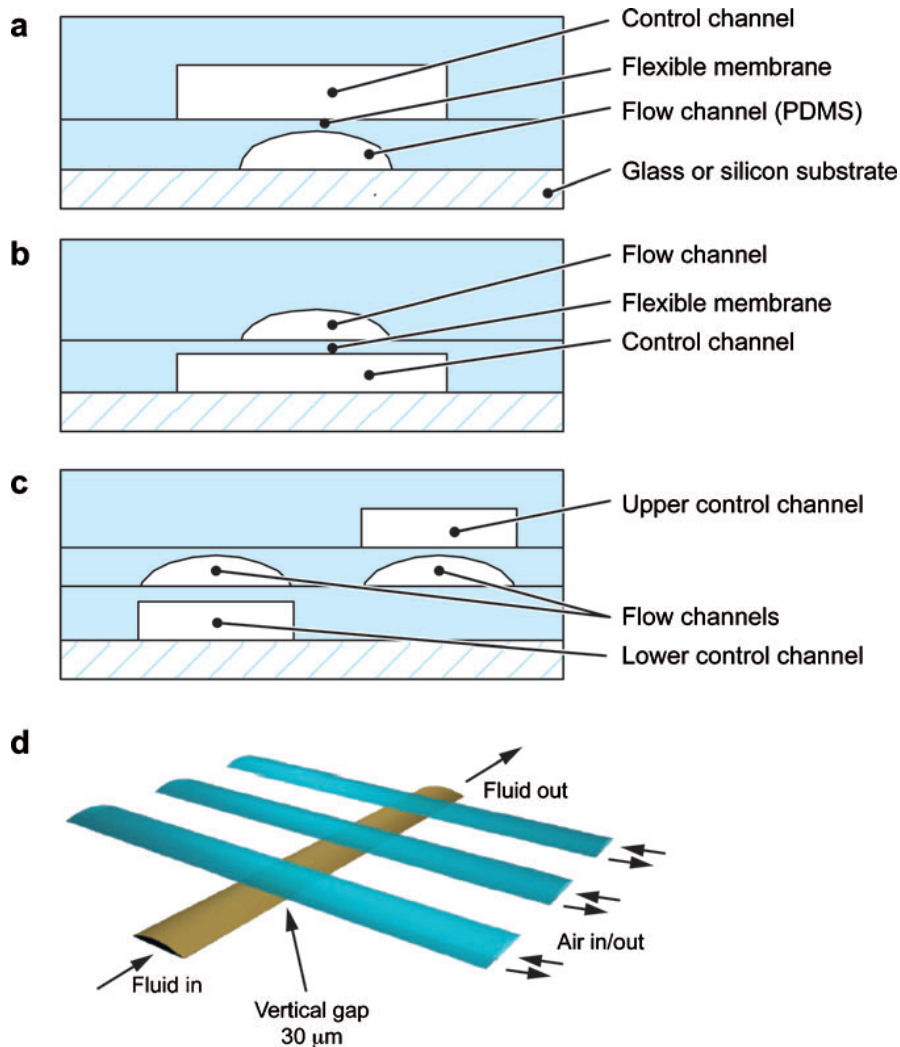
5. Microfluidics

4. Microtransfer molding  
( $\mu$ TM)

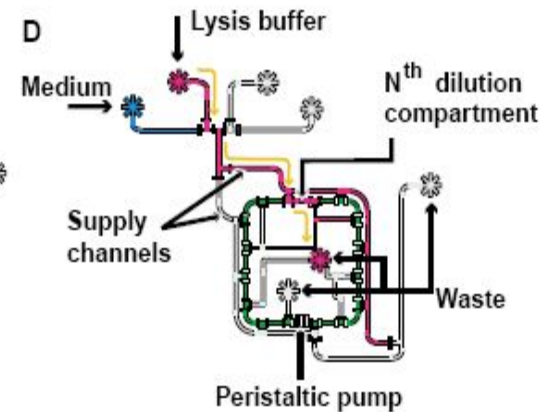
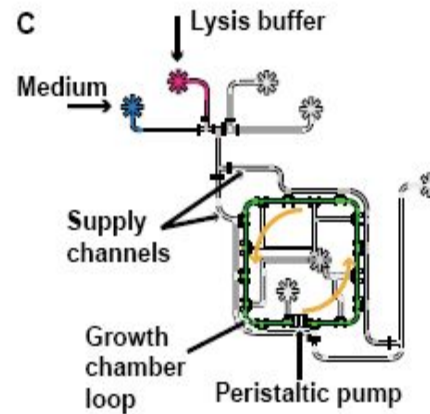
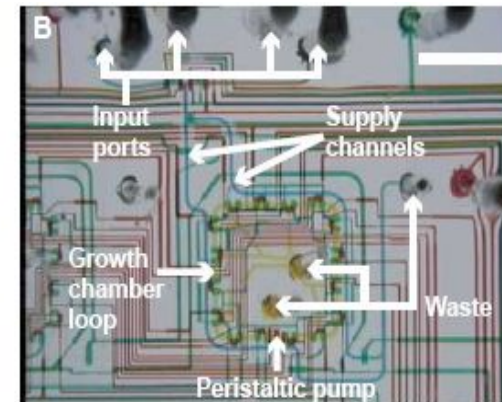
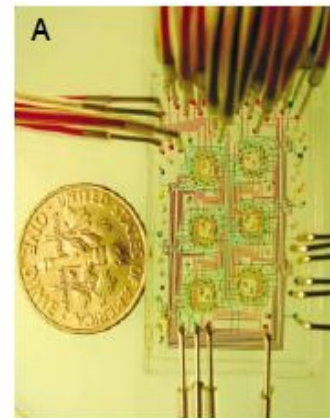


# & Multilayer Soft Lithography

...From Simple Valving...

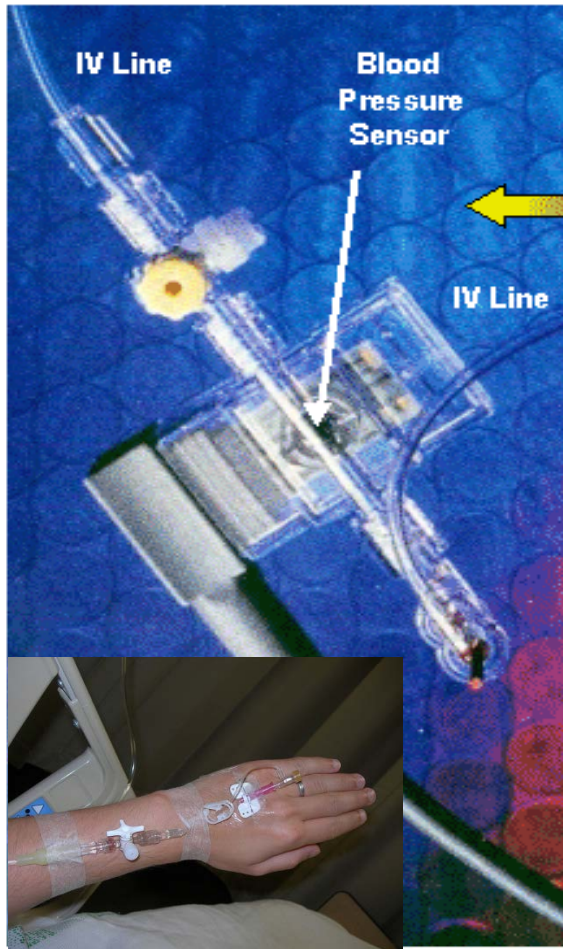


...to Complex Systems:  
A microfluidic Chemostat

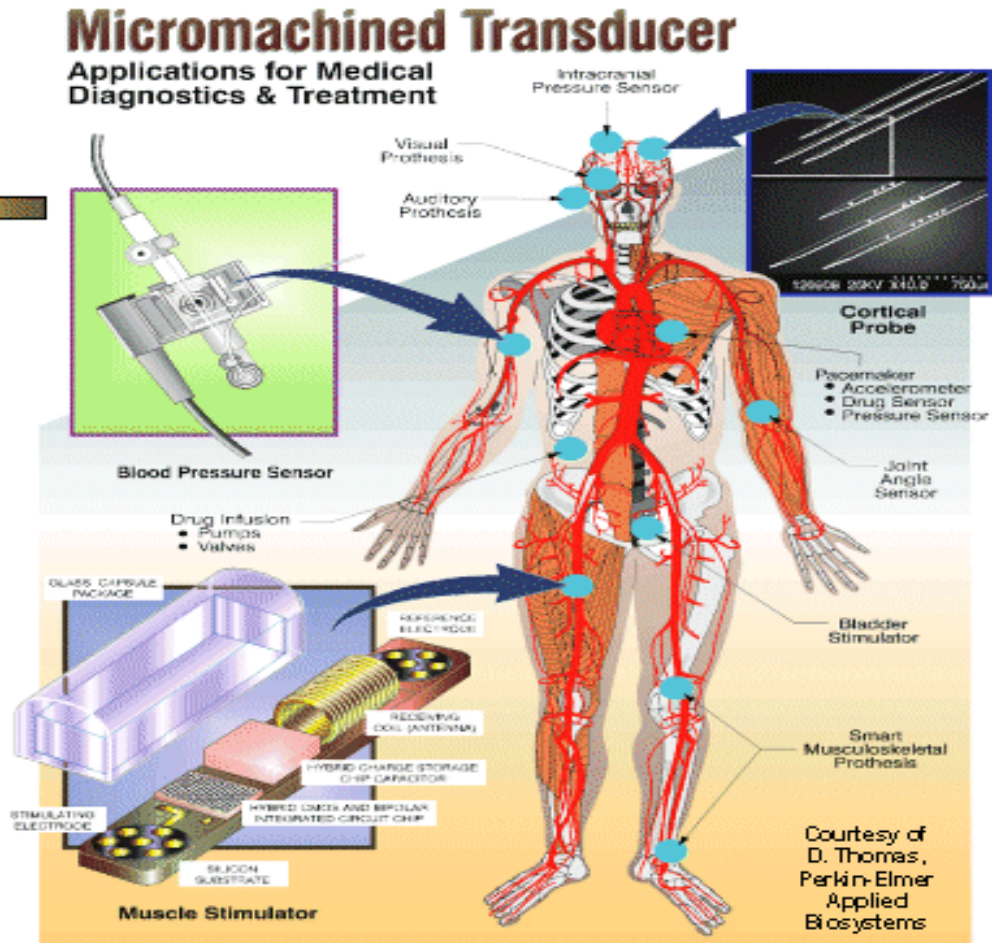


# BioMEMS in the Medical Field

Ex vivo...

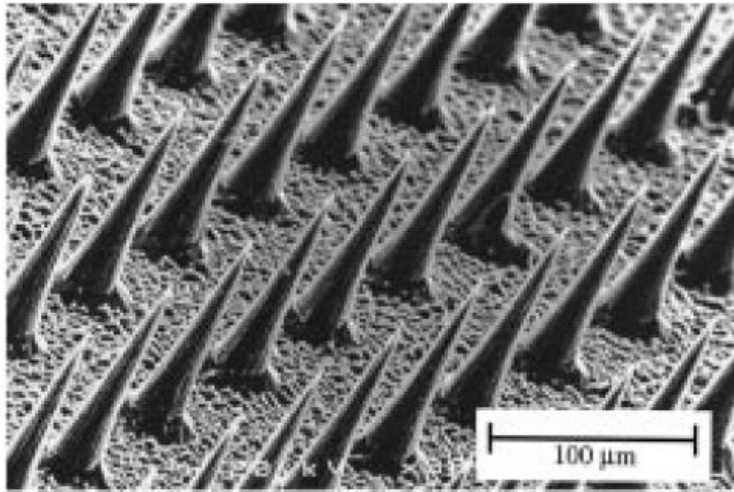


In vivo...

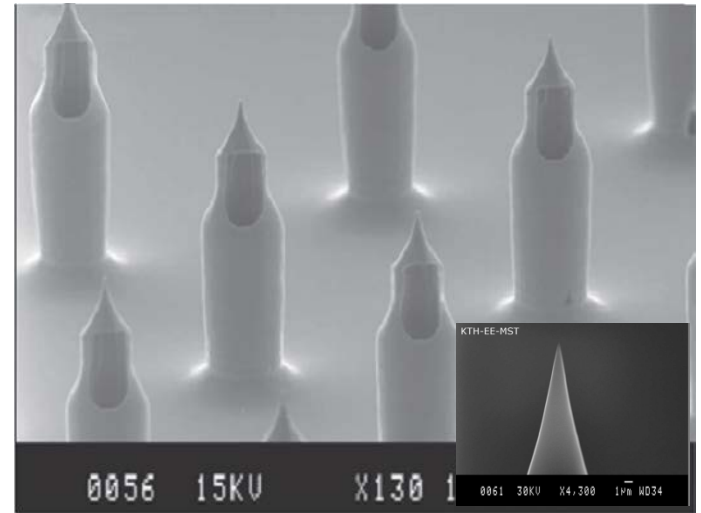


# Micro Needles

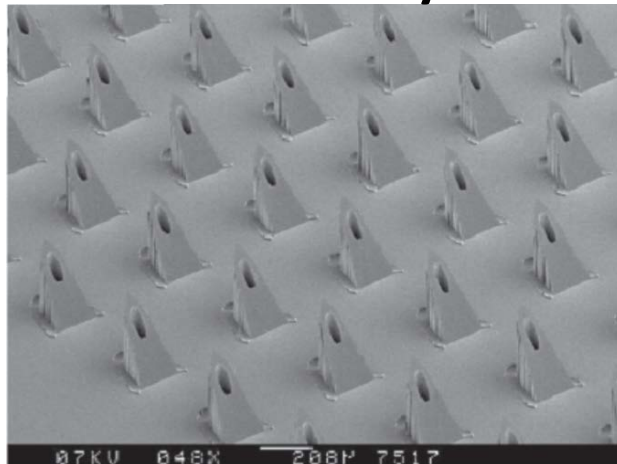
**Solid MicroNeedles  
(coated, first generation)**



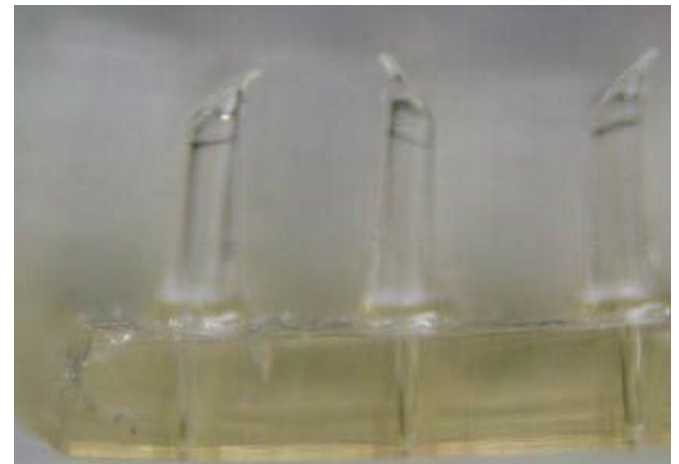
**Ultrasharp Si (Citadel style)  
with a hole at the side**



**Saw-tooth style**



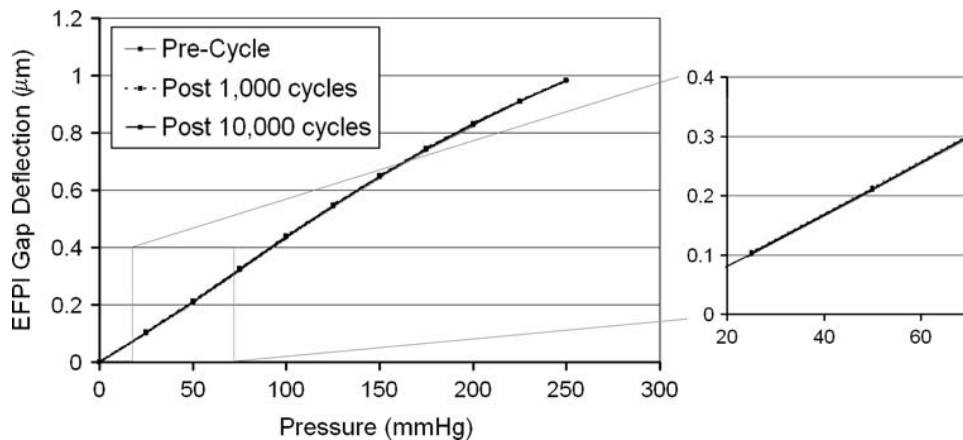
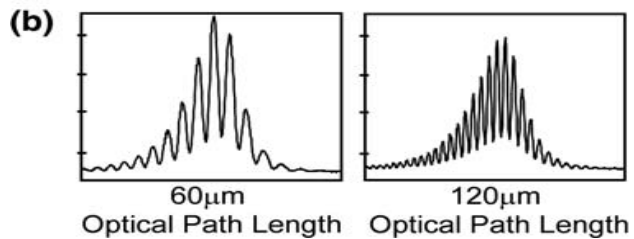
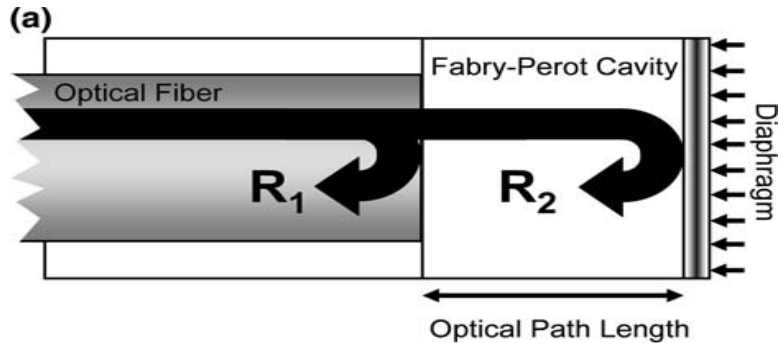
**Polymer-based (PDMS)**



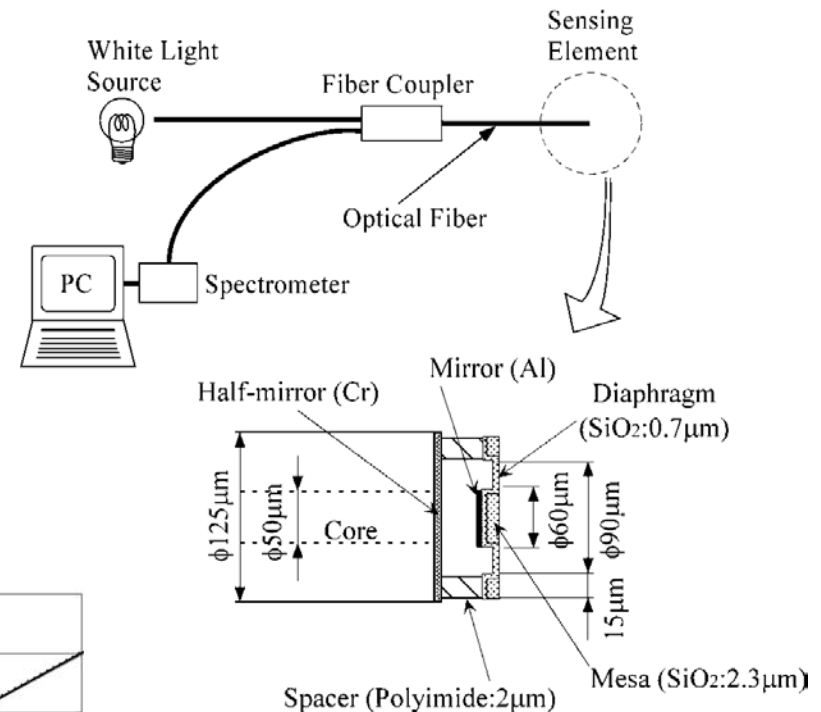


# Optical Pressure Sensors

**Concept:** A deformable membrane acts as a mirror in a Fabry-Pérot cavity



## The Measuring Setup



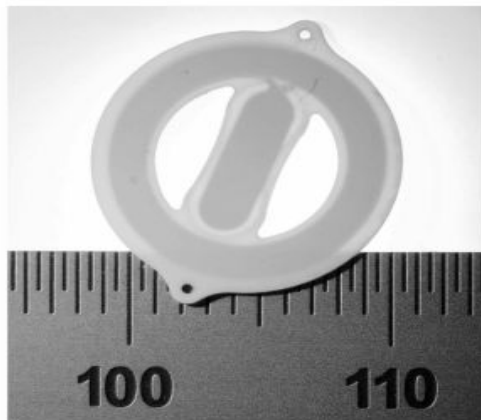
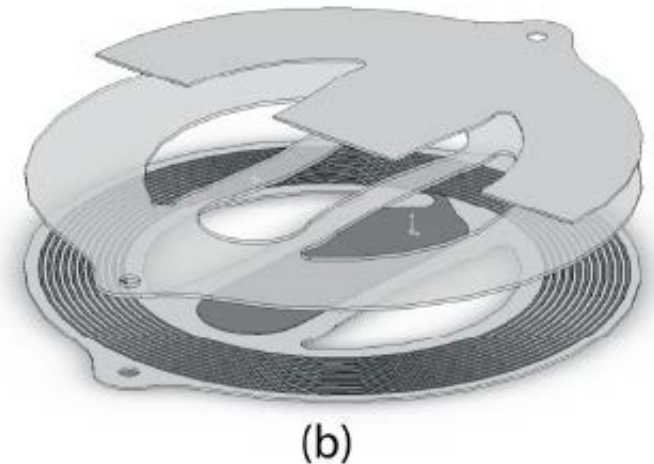
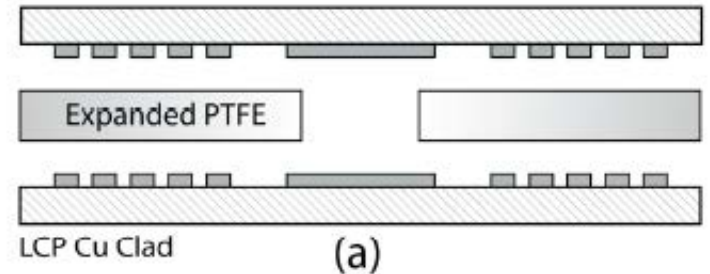
# The CardioMEMS Sensor

## Materials

- Copper-clad Liquid Crystal Polymer (LCP)
- Expanded polytetrafluoroethylene (PTFE)

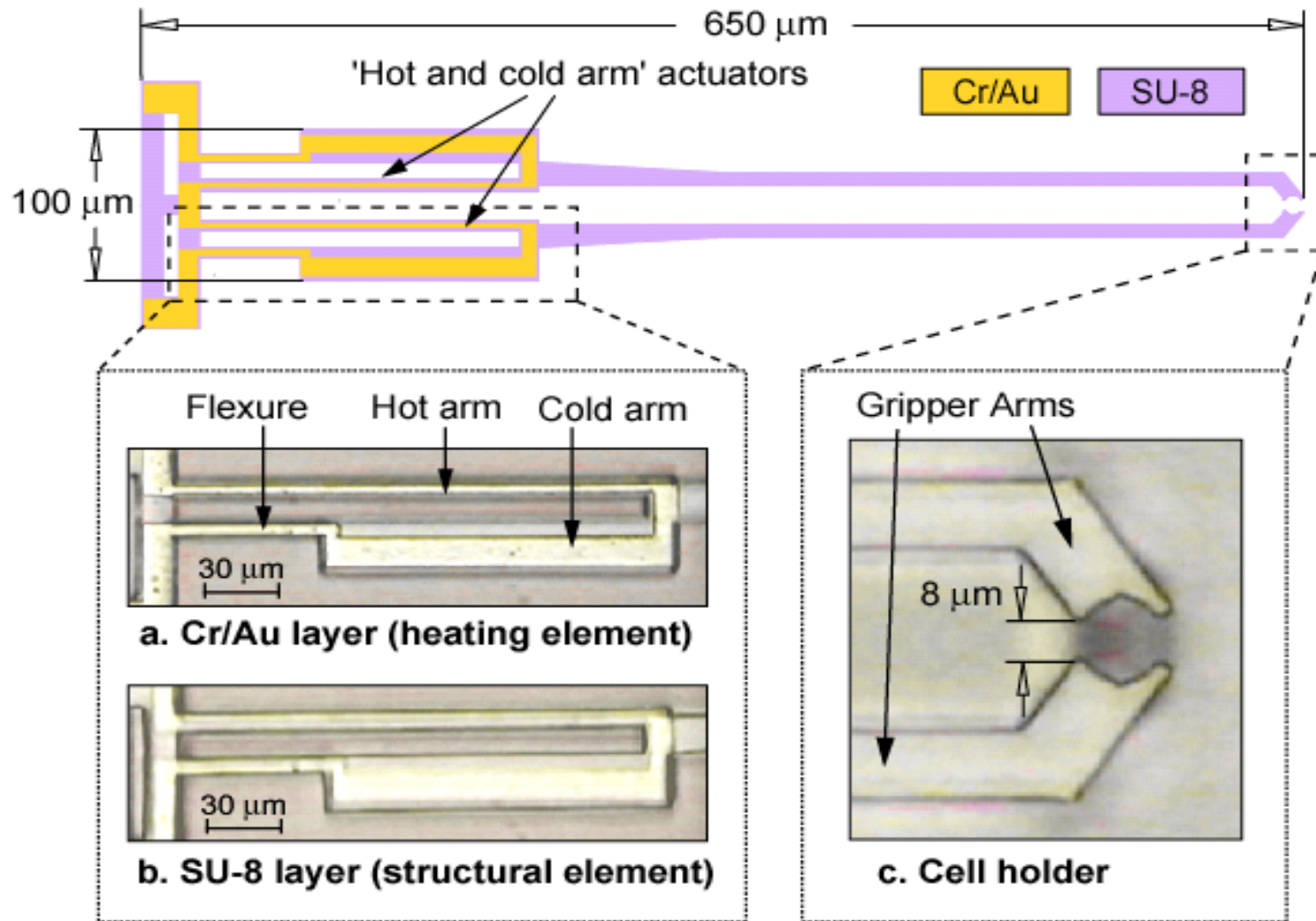
## Microfabrication Process

- Photolithography/ Wet Etching
- Bonding: The layers are aligned, assembled and laminated at 180°C under pressure

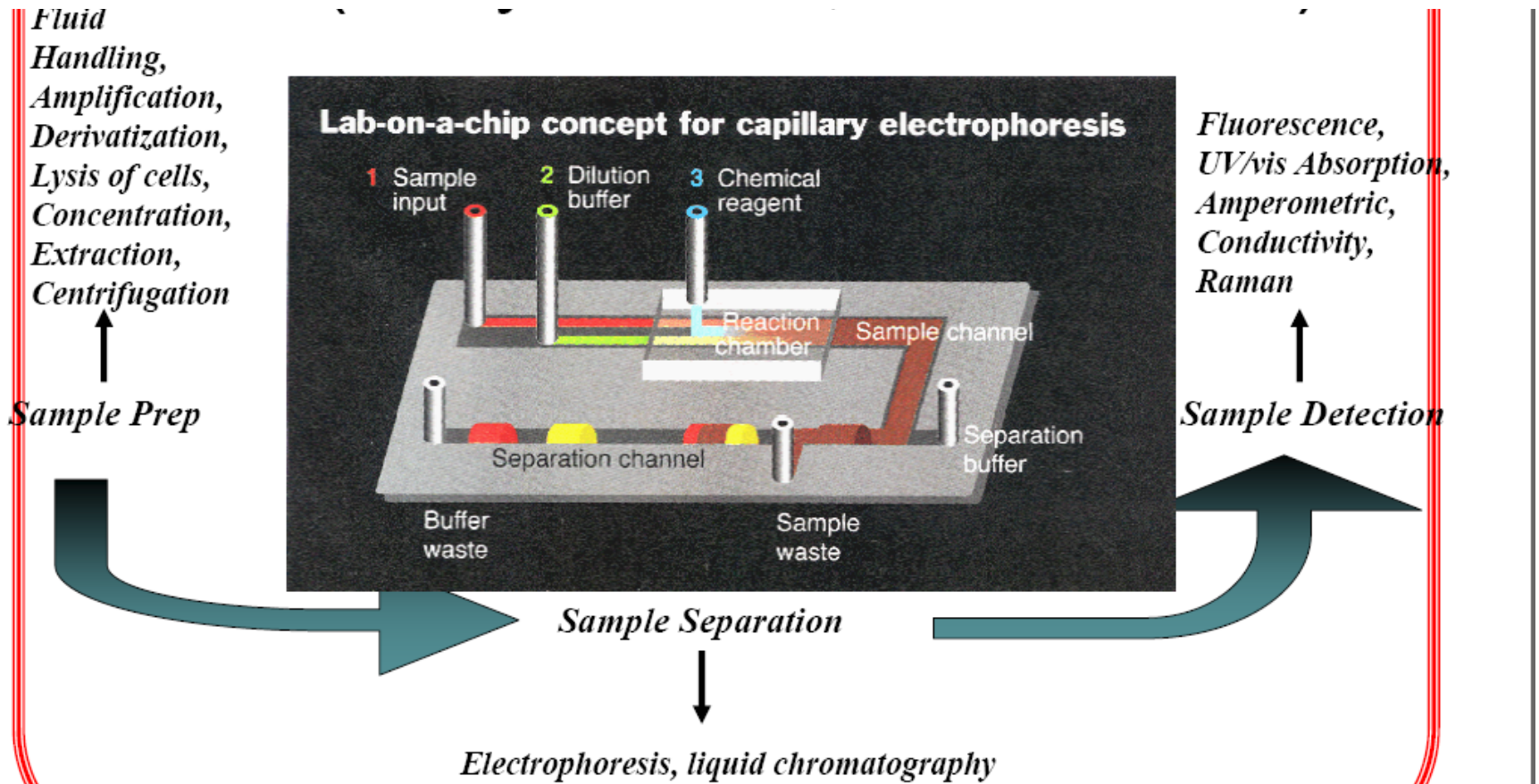


**Final Device:** A self-packaged structure in which only a polymer outer surface is exposed to the environment

# BioMEMS Actuators



# Microfluidics/Lab-on-Chip Systems



# Navier-Stokes Equations

$$\rho \left( \frac{\partial \vec{V}}{\partial t} + \vec{V} \cdot \nabla \vec{V} \right) = -\nabla p + \rho \vec{g} + \mu \nabla^2 \vec{V}$$

The diagram illustrates the decomposition of the Navier-Stokes equation. A horizontal line is drawn under the left-hand side of the equation, with a blue arrow pointing down to the text "Inertial Forces". Another horizontal line is drawn under the  $-\nabla p$  term, with a blue arrow pointing down to the text "Pressure Forces". A third horizontal line is drawn under the  $\rho \vec{g}$  term, with a blue arrow pointing down to the text "Gravity Forces". A fourth horizontal line is drawn under the  $\mu \nabla^2 \vec{V}$  term, with a blue arrow pointing down to the text "Viscous Forces".

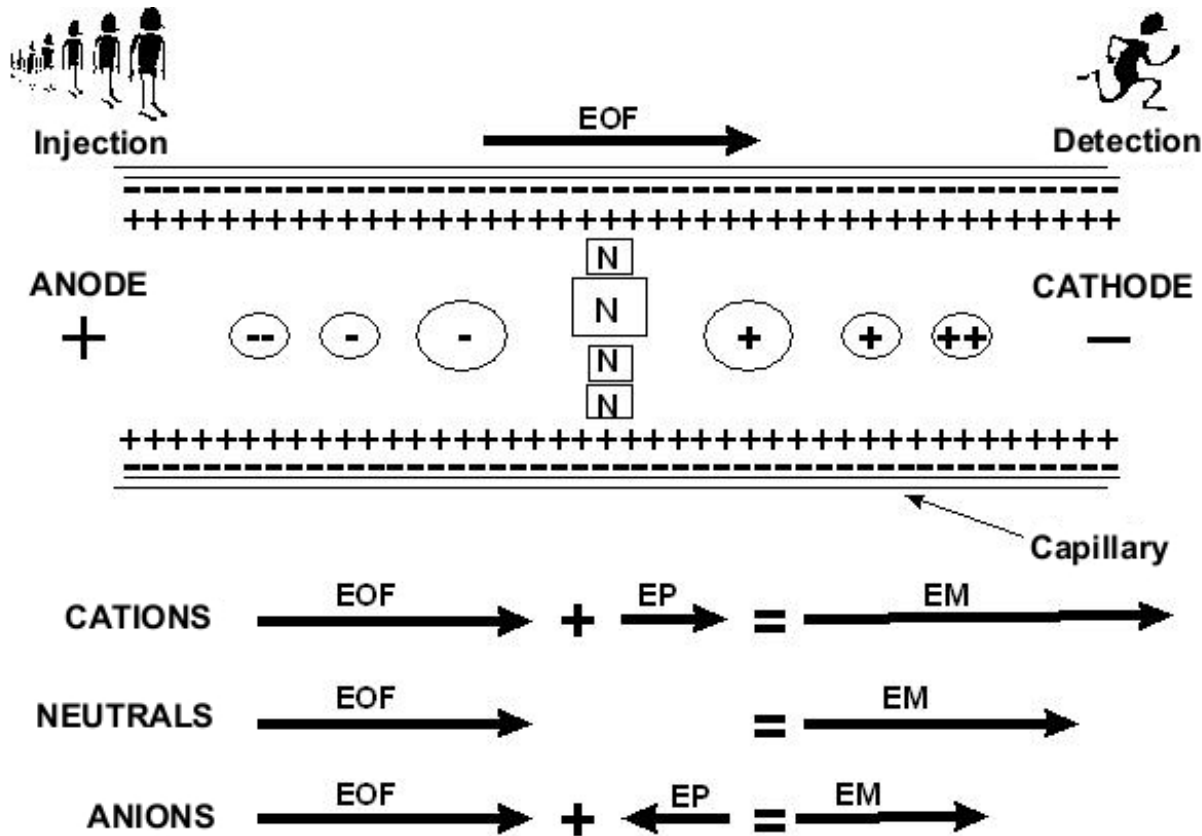
In most microfluidic cases, Inertial & Gravity forces are negligible compared to Pressure & Viscous forces

$$\text{N-S: } 0 = -\nabla p + \mu \nabla^2 \vec{V}$$



# EOF and Electrophoresis

*EOF and Electrophoresis might compete each other...*



*Do not forget to calculate absolute velocities:*

$$\vec{u}_{abs} = \vec{u}_{ep} + \vec{u}_{EOF}$$

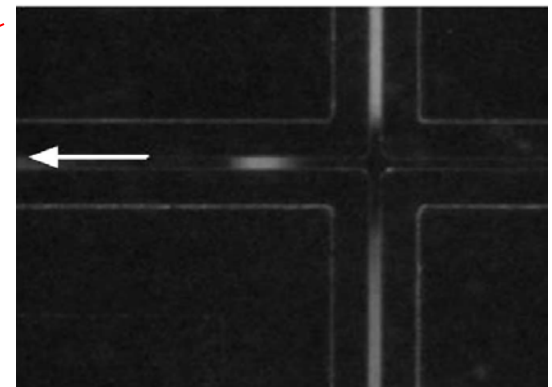
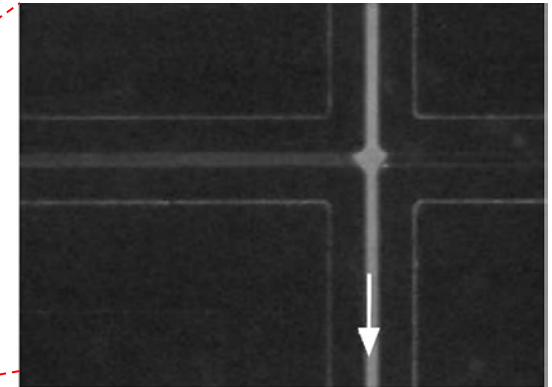
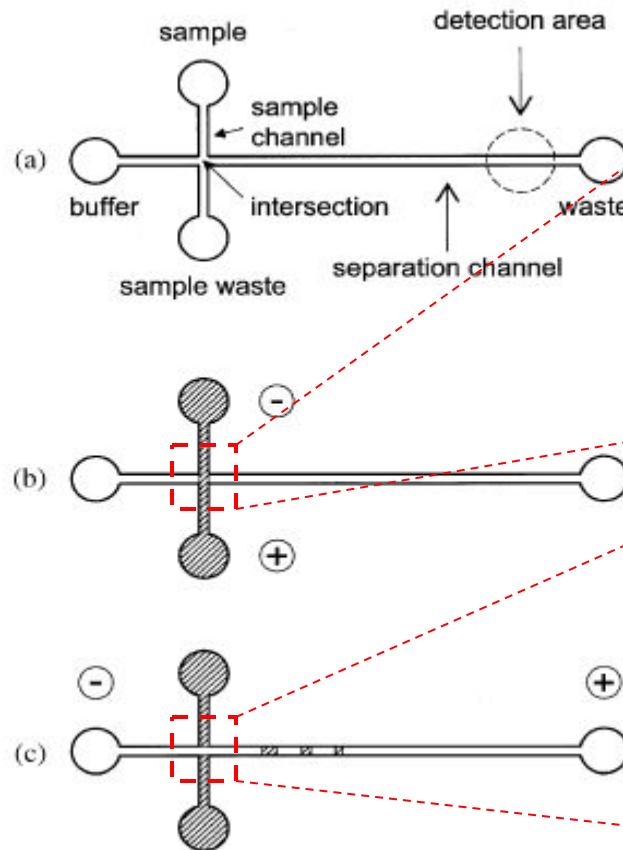
# Capillary Electrophoresis for DNA Separation

Concept:

Use microfluidic channels (capillaries) to separate DNA fragments

## Operation Principle

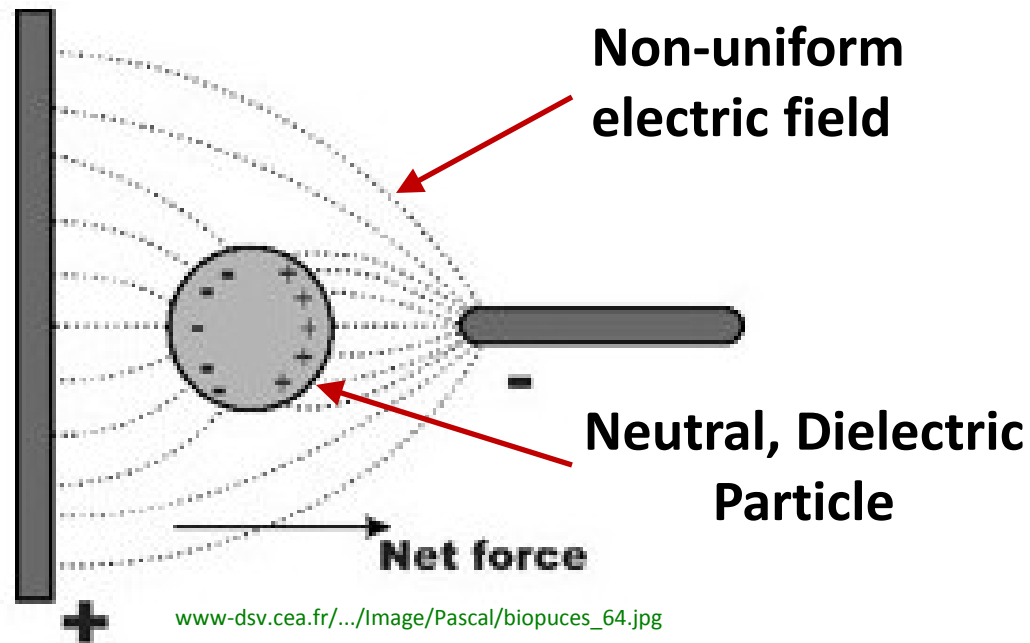
- Fill the channel intersection with sample solution
- Apply potential between buffer and waste inlets to initiate electrophoresis



**Electric Field applied: 200-400 V/cm, Separation time: 1-2 min, Limiting factor: Joule Heating**

# Dielectrophoresis

An Non-uniform Electric Field exerts a force on a uncharged, dielectric object (e.g. particle)



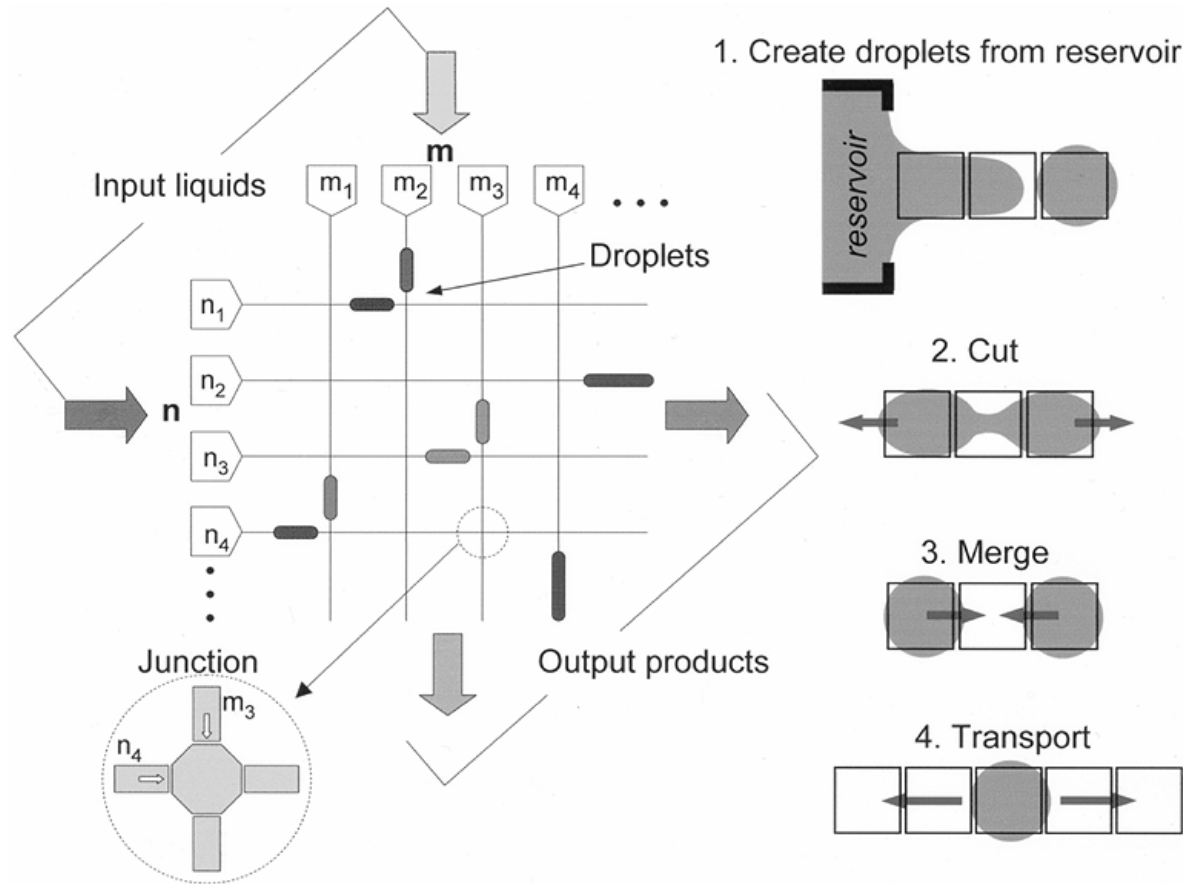
The object does not have to be charged,

All dielectric objects exhibit dielectrophoretic activity!

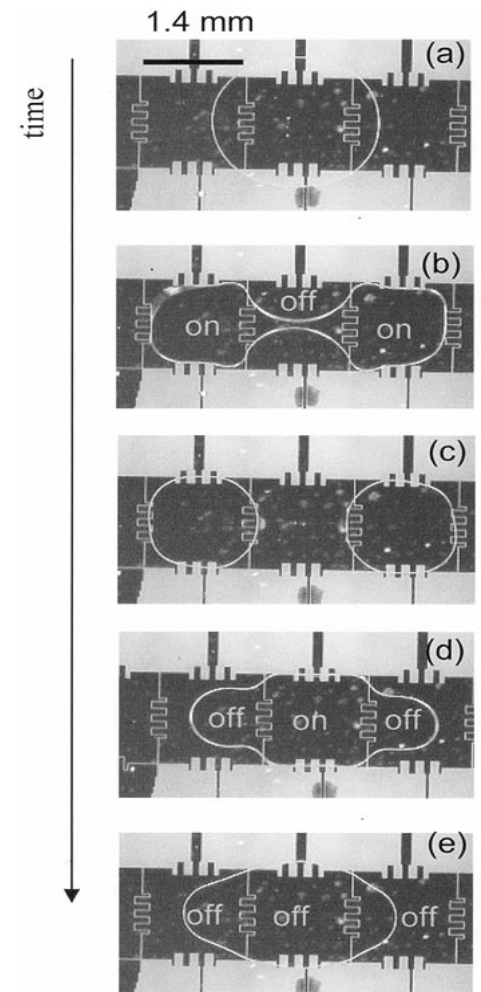
## Application

*To move, trap, separate, neutral, dielectric objects (e.g. cells)*

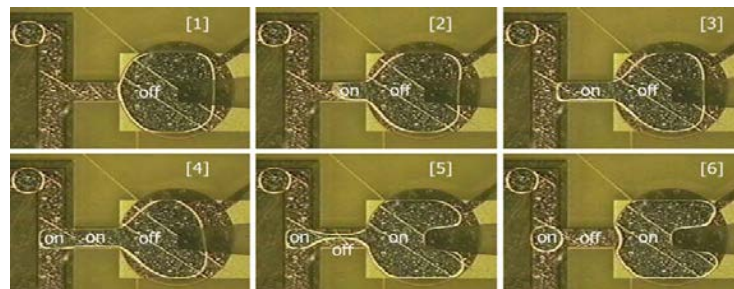
# Fluidic Operations in Digital $\mu$ fluidics



## 2. Cut & Merge (Split & Mixing)



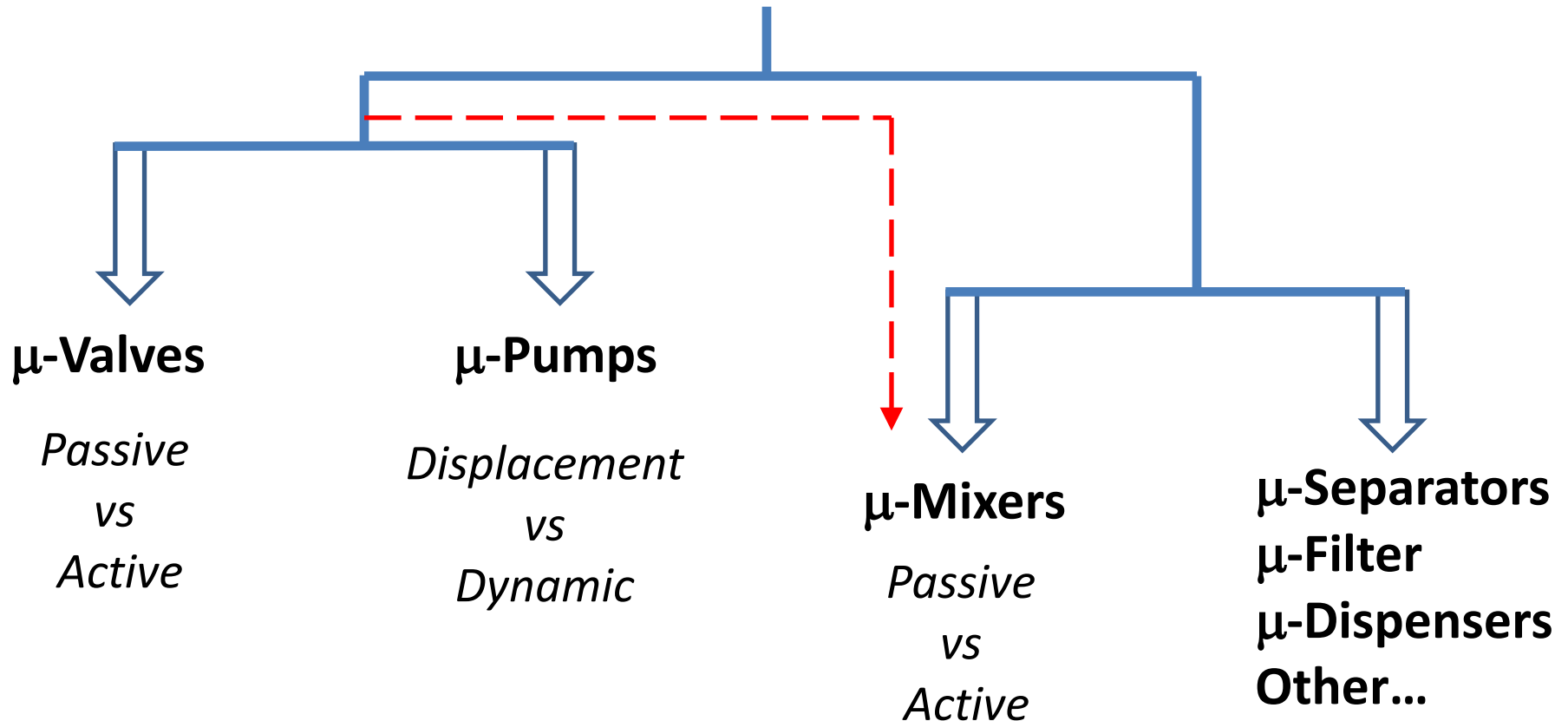
## 1. Droplet formation/injection



# Microfluidic Components



Used to manipulate  
(transport, mix, separate, etc) fluids

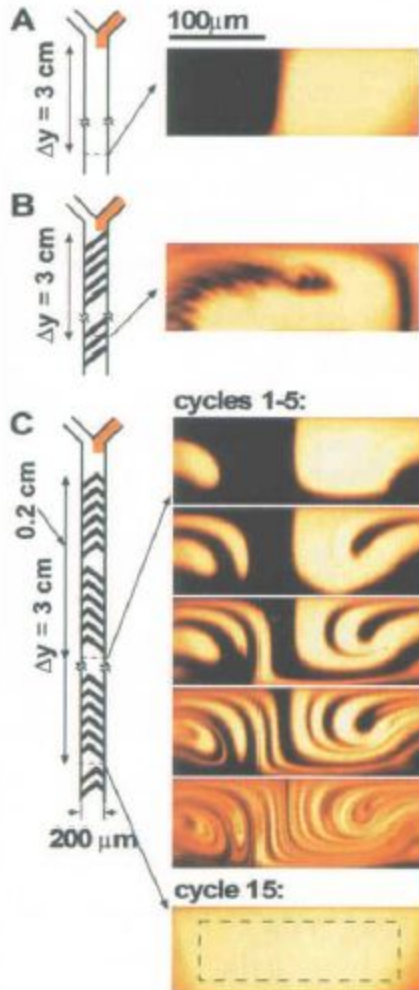


# The Herringbone Mixer

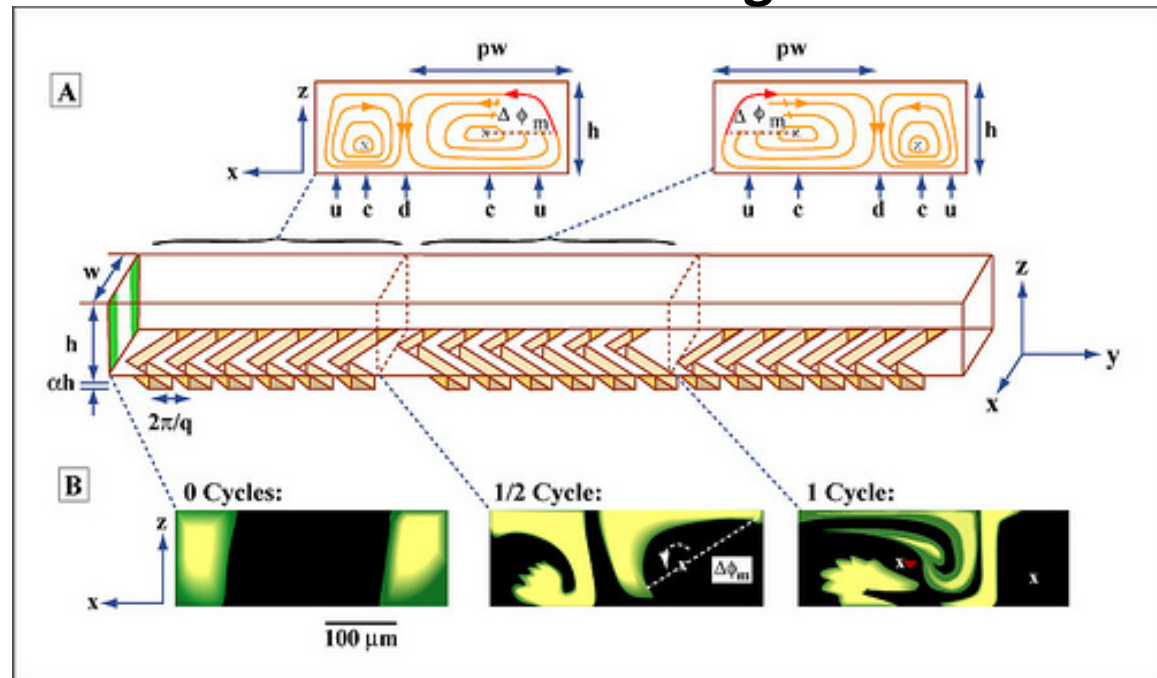
## Concept:

Use set of ridges to create transverse vortices, (parallel to the cross section of the channel)

## 2-Flow Mixing



## 3-Flow Mixing



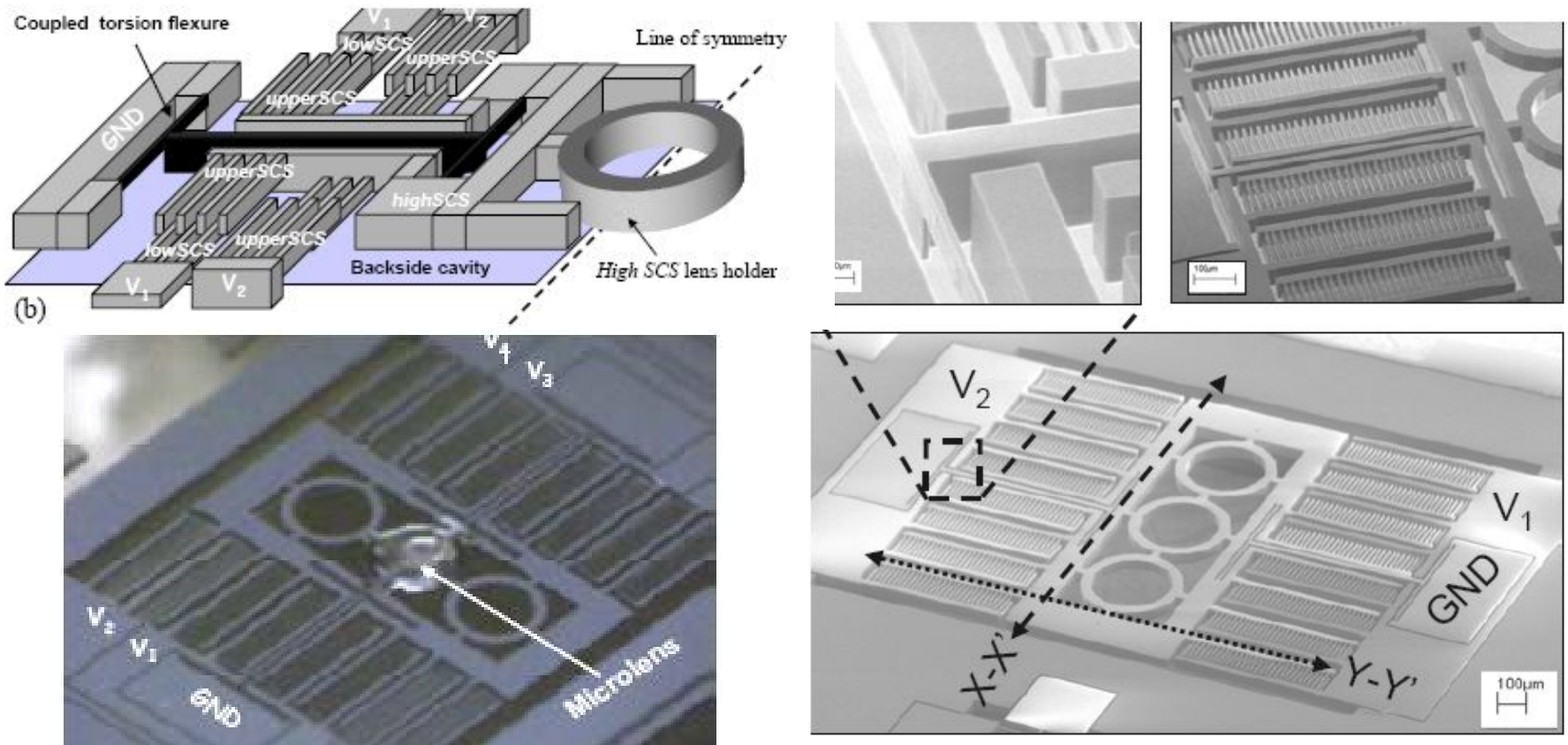
- Channel Width = 200  $\mu\text{m}$ , Channel Height = 70  $\mu\text{m}$ , Ridge Depth = 40  $\mu\text{m}$ , Ridge Width ridge = 200  $\mu\text{m}$
- Mixing length 1-3 cm,  $\text{Re} \sim 10^{-2}$



# Integration. $\mu$ -lenses on $\mu$ -Actuators

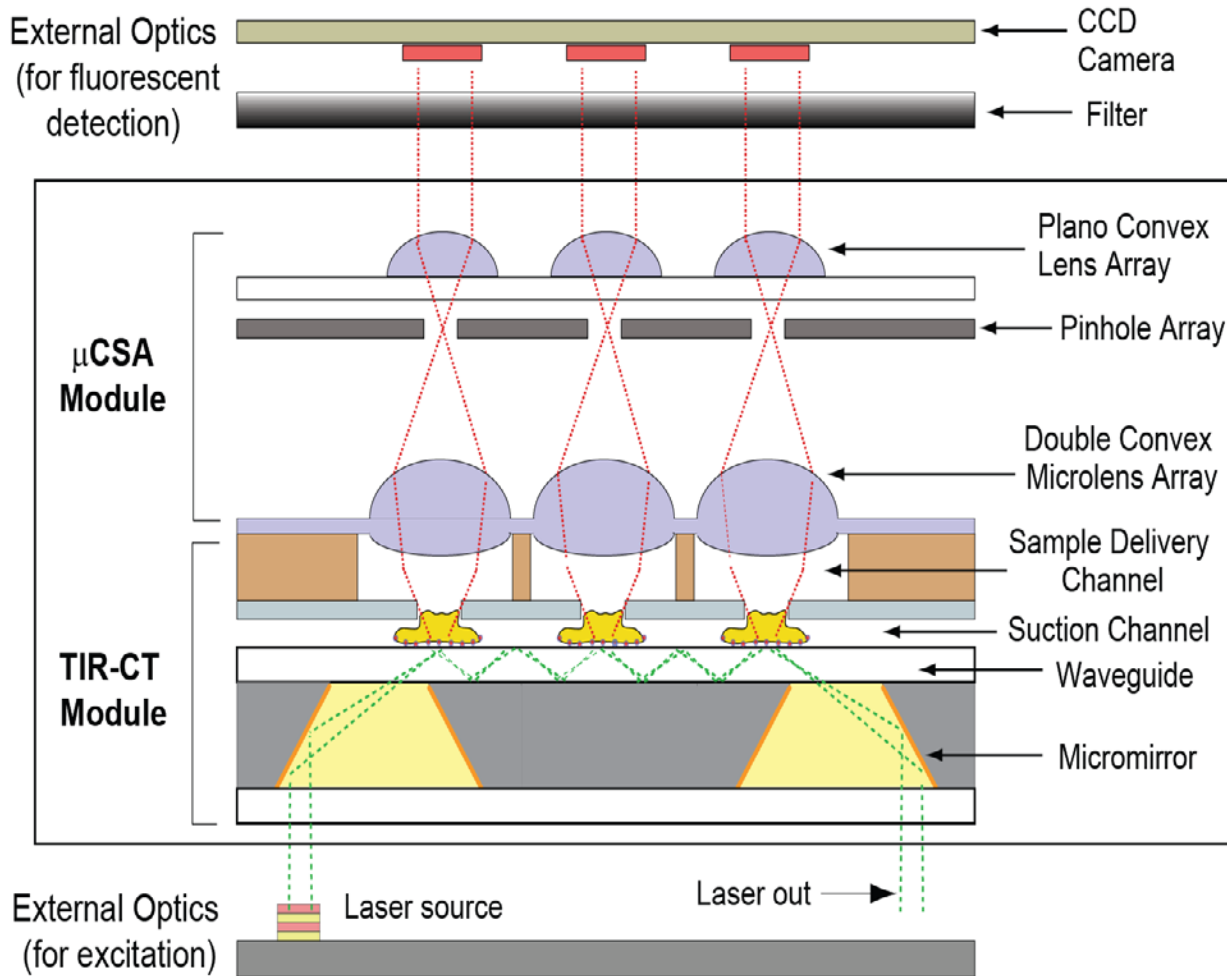
## Concept

*Integrate electrostatic  $\mu$ -actuators with  $\mu$ -lenses (e.g. for scanning...)*



- $\mu$ -lenses are simply dispensed on the actuator ring and UV cured...
- Electrostatic actuators (comb drives) are used as they require minimum power

# Integration. Optical Detection and Excitation on-chip



The biochip integrates two modules:

- the **TIR-CT module** for Isolating, Trapping and Illuminating single WBCs
- the **μCSA module** for imaging/counting the trapped WBCs



**Some other exciting stories...**

# 1. Single Molecule Real Time (SMRT) Sequencing

**Motivation: The \$1,000 Genome Project**

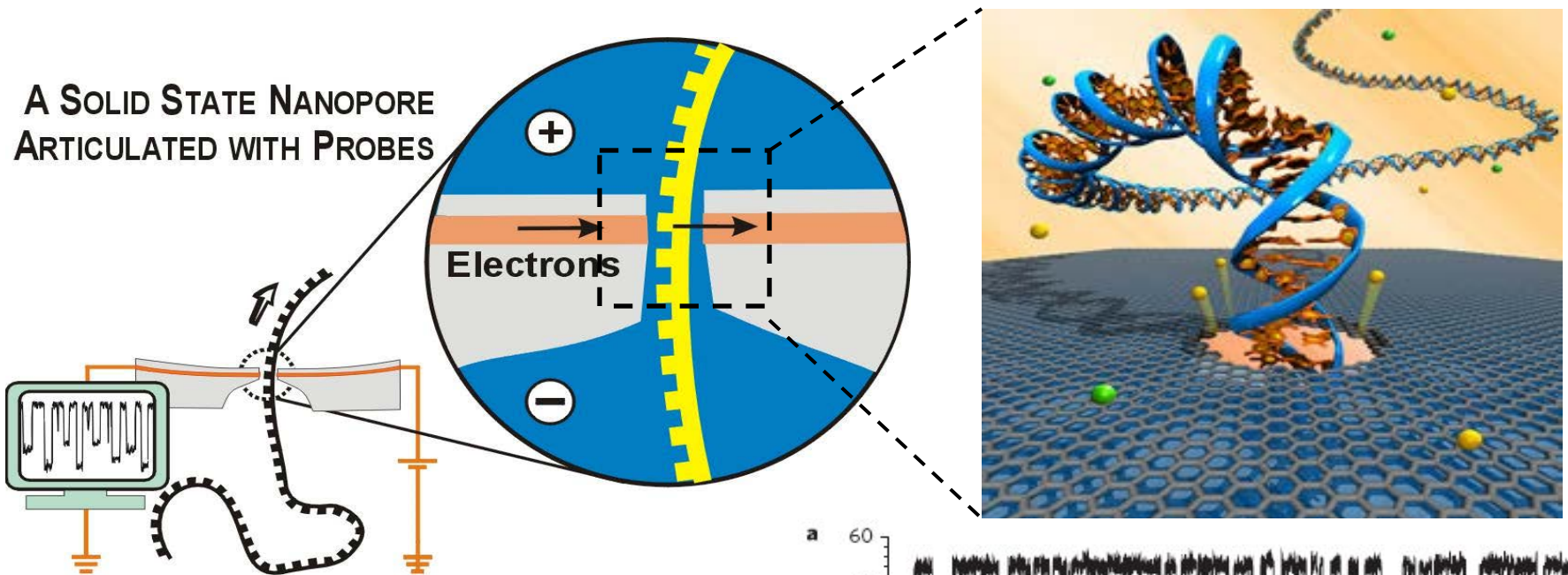
What if you could sequence the entire human genome in a single day, in a single experiment — for less than \$1,000?

# Nanopores for DNA SMRT Sequencing

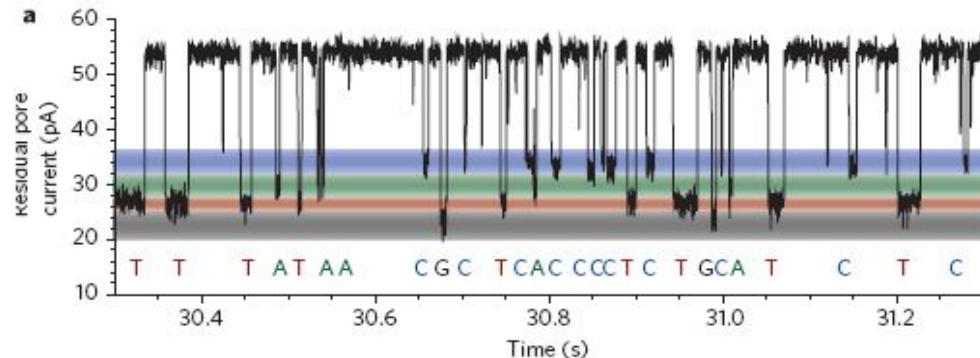
## Concept

*Flow DNA through a (~1nm) nanopore and measure the electric current*

A SOLID STATE NANOPORE  
ARTICULATED WITH PROBES

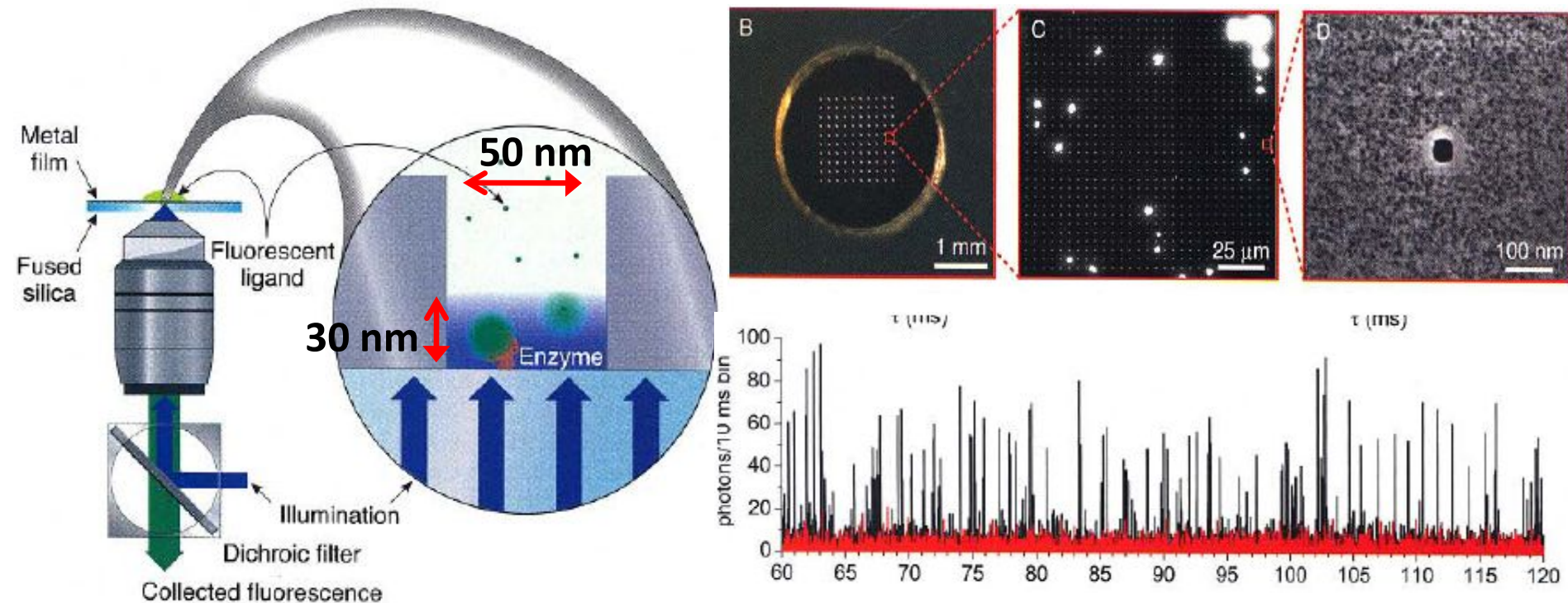


*...The amount of current which can pass through the nanopore at any given moment depends on whether the nanopore is blocked by an A, a C, a G or a T*



Currently under development by several companies (Oxford Nanopore Technologies, Noblegen)

# Zero-mode Waveguides for DNA SMRT Sequencing



*Zero-mode waveguides (ZMW) guides light into a volume that is small in all dimensions compared to the wavelength of the light:*

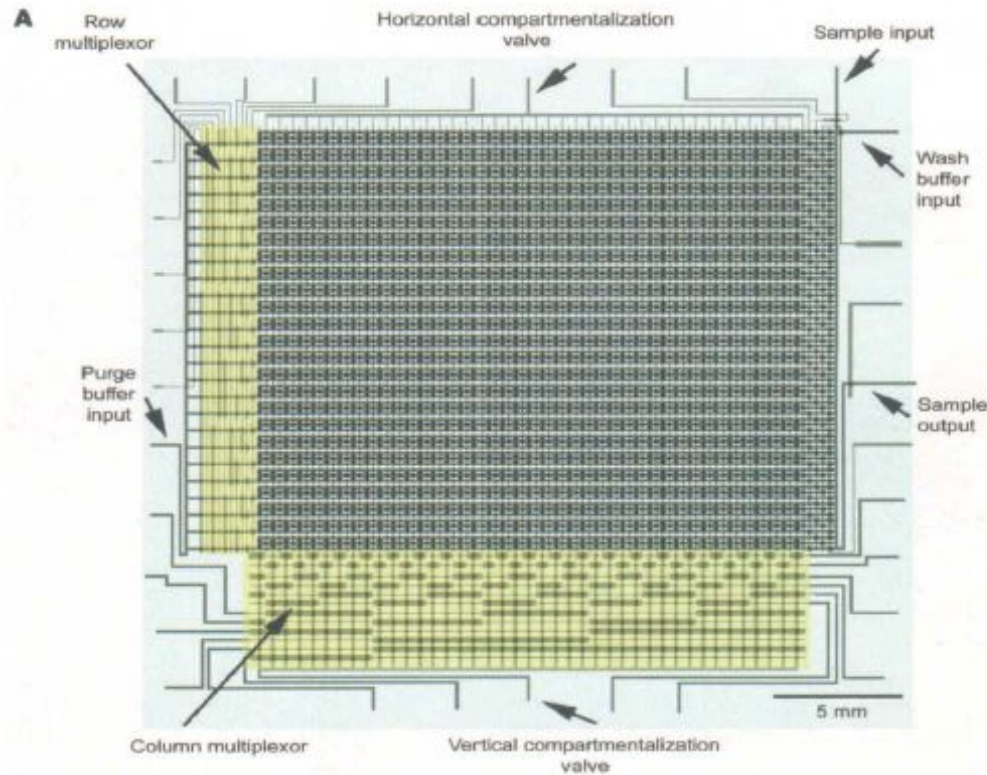
**→ Minimize background noise → Single Molecule Imaging**

*Under development by Pacific Biosciences*

## **2. Large Scale Microfluidic Handling**



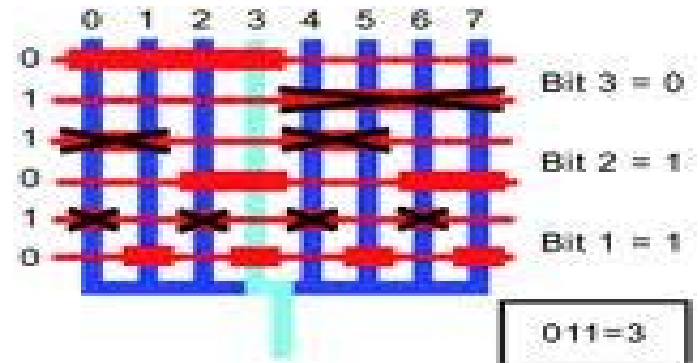
# Large-Scale Integration of $\mu$ -valves



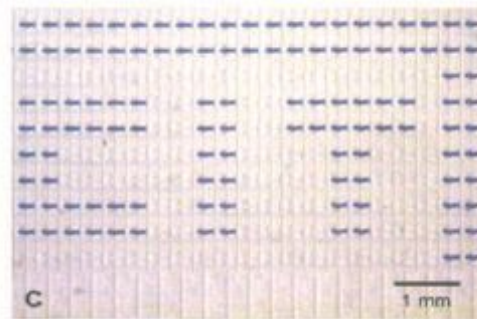
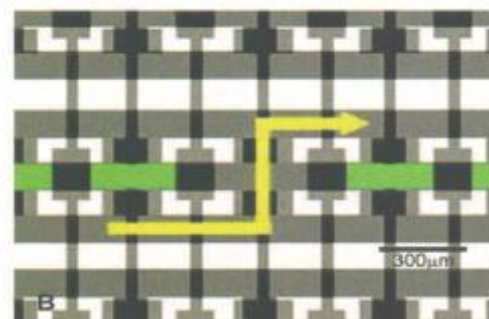
## SPECS

- 3574 on-chip  $\mu$ -valves
- 22 outside control interconnects
- 1,000 individually addressable picoliter reaction chambers
- A column and row multiplexor are used to address each chamber

## The microfluidic Multiplexor



Reference: 'Microfluidic Large-Scale Integration',  
*Science*, 2002, Vol. 298 no. 5593 pp. 580-584



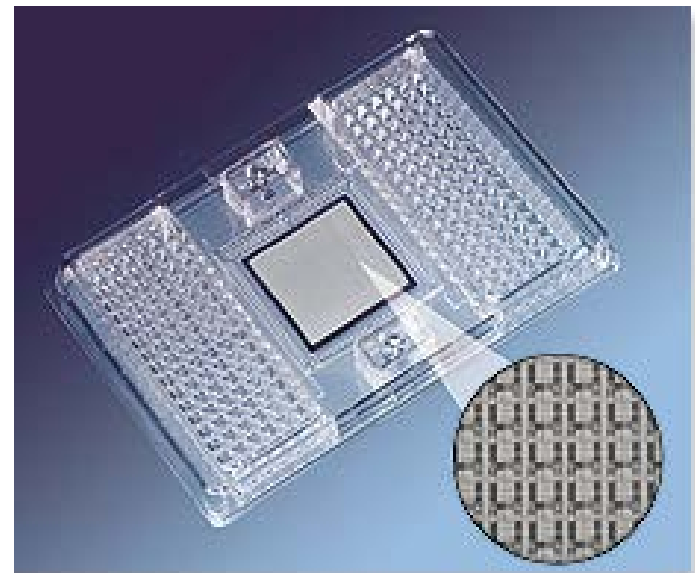
# Fluidigm Dynamic Array Integrated Fluidic Circuits (IFCs)

## On-chip High-throughput Polymerase Chain Reaction (PCR)

*Fluidigm chips have an on-chip network of microfluidic channels, chambers, and valves that automatically assemble up to 2,304 unique PCR reactions, decreasing the number of pipetting steps required by up to 100 fold.*

### Applications

- Gene Expression
- SNP Genotyping
- Targeted Resequencing
- Single-Cell Gene Expression
- Protein Crystallization
- ...



Watch Videos at: <http://www.fluidigm.com/biomark-videos.html>

# 3. Centrifugal Microfluidics

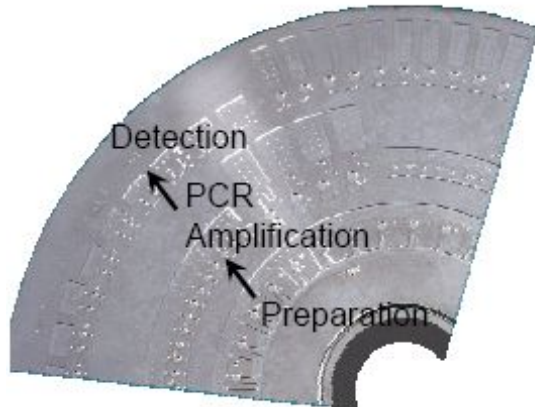
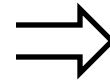
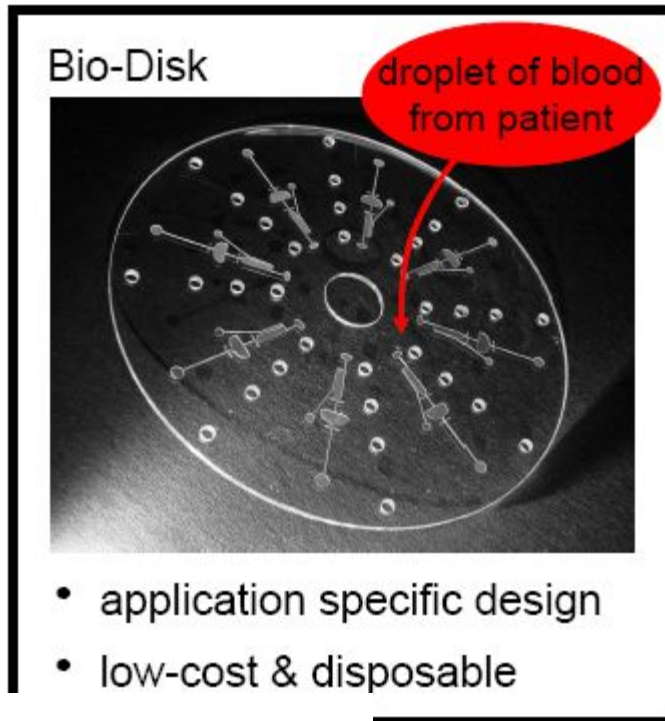


Commercialized by GYROS:

[http://www.gyros.com/en/company/about\\_gyros/index.html](http://www.gyros.com/en/company/about_gyros/index.html)



# The GYROS BioDisk

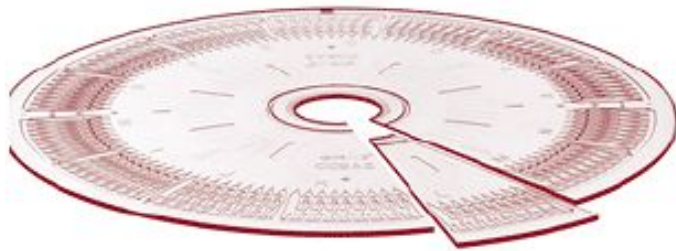


## Key Idea:

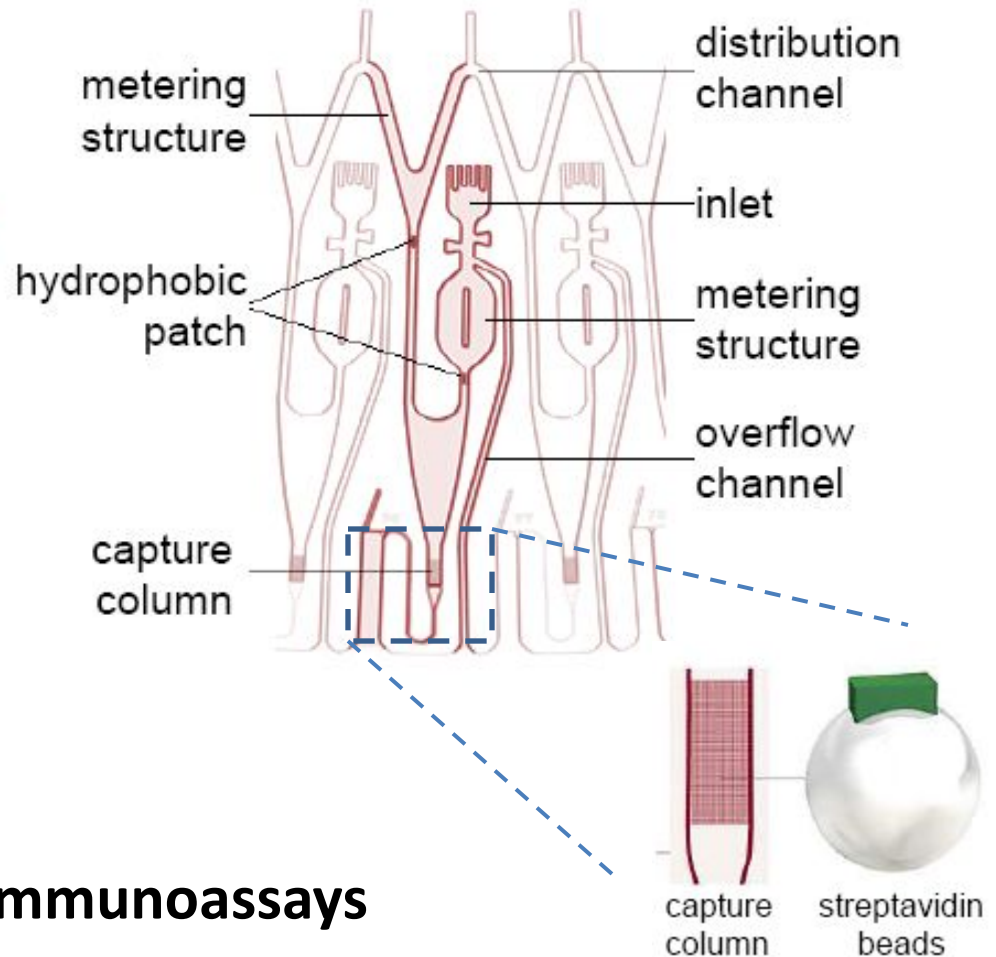
*Use hydrophobic Patches to block fluid flow. Use Centrifugal Forces to overcome these pads*

# GYROS for Protein Quantification

CD for protein quantification



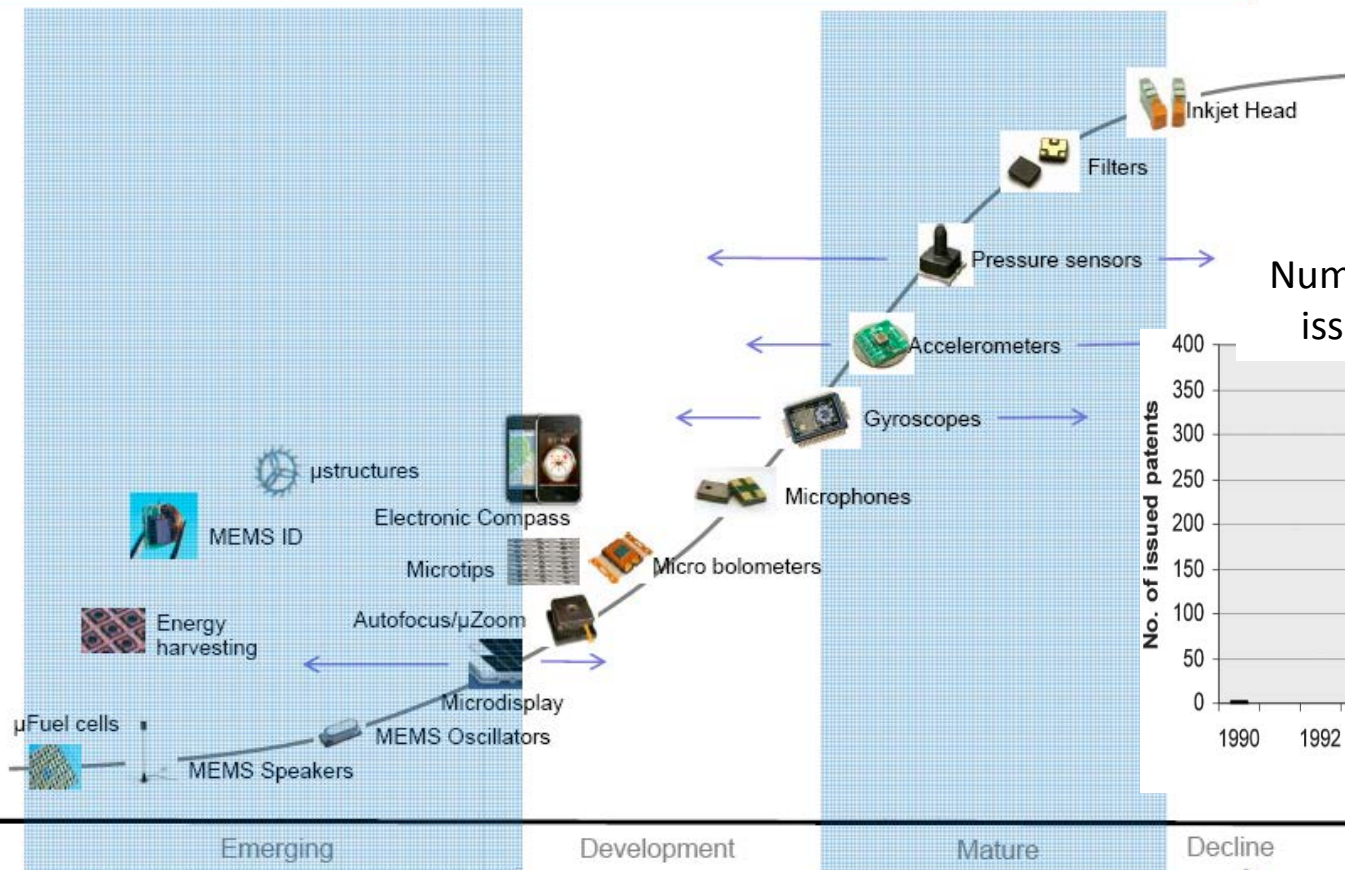
- 112 parallel measuring structures per CD
- 200 nL of sample and reagent per measurement
- time-to-result < 1h



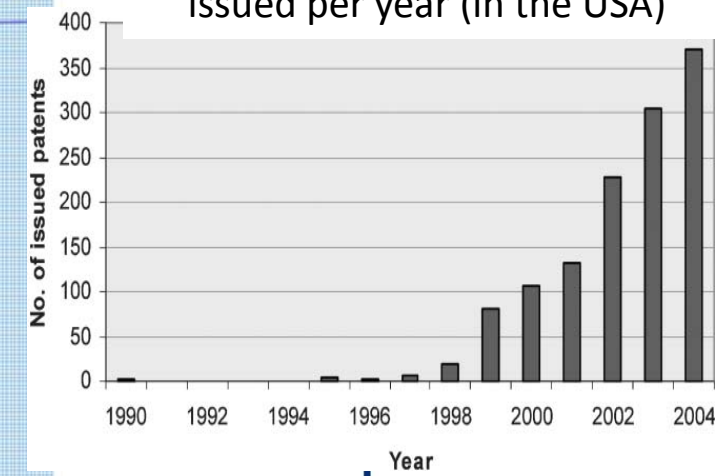
**Applications: Point-of-Care Immunoassays**

# BioMEMS: The future is Bright!

## MEMS products phase of development



Number of microfluidic patents issued per year (in the USA)



**Hope you got Inspired!**

*...And please do not forget to evaluate the class...*